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# The Effectiveness of Insect Growth Regulator (IGR) on the Growth and the Development of *Aedes aegypti* and *Aedes albopictus* in Tangerang City, Indonesia

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#### Abstract

**Background**: Dengue fever is a world health problem. It is endemic in more than 100 countries and threatens about 2.5 billion or 40% of people living in urban, suburban, and rural areas in both tropical and subtropical climates. Various mitigation measures have been undertaken, including vector control using mosquito nest eradication, selective abatization with larvicides, and mosquito fogging. Elimination using chemicals causes many problems. Another alternative to control the growth and development of Aedes sp. mosquito is needed. The use of insect growth regulator (IGR) is suggested to overcome this problem, since it does not affect the cleanliness of water, does not increase resistance, and considered safe.

**The Objective**: The objective of this research was the  $IE_{50}$ ,  $IE_{90}$ , and  $LC_{50}$  concentration of pyriproxyfen to eliminate Aedes aegypti and Aedes albopictus larvae.

**Method**: The type of research used was laboratory experimental with a post-test design with only one group control design. This research was done through the actual test by creating dose variations of 1000 ppm of pyriproxyfen, which were as much as 0,14 mL; 0.196 mL; 0.274 mL; 0.384 mL; 0.538 mL; and 0.752 mL. The results of observation of larvae mortality were calculated by probit analysis to obtain the  $LC_{50}$  value of pyriproxyfen against Aedes aegypti and Aedes albopictus. The  $LC_{50}$  value obtained from probit calculation was replicated four times and used to find the value of  $LT_{50}$ .

**Results**: Aedes aegypti and Aedes albopictus larvae after pyriproxyfen treatment and probit calculation showed inhibition of the growth Aedes aegypti larvae, indicated by  $IE_{50}$  value (95% Fiducial limits (FL)) of 0.17ppb (0.13-0.21) and  $IE_{90}$  (95% FL) of 0.68ppb (0.43-1.08). Inhibition of the growth of Aedes albopictus larvae was indicated by the value of  $IE_{50}$  (95% FL) of 0.11ppb (0.09-0.13) and  $IE_{90}$  (95% FL) of 0.39ppb (0.26-0.57). External morphological changes also occurred to Aedes aegypti and Aedes albopictus larvae after the exposure. Flaking of the cuticle to the thorax and swelling of both the head and the thorax were observed. Therefore, insect growth regulator has the potential to prevent the development of larvae into adult mosquitoes.

**Conclusions**: Pyriproxyfen can inhibit the growth of larvae. Pyriproxyfen  $LC_{50}$  concentration on Aedes aegypti larvae were 1.63ppm and on Aedes albopictus larvae was 1.56ppm. The use of pyriproxyfen causes external morphological changes to occur in larvae, pupae, and adult mosquitoes especially on the feet, whereas the proboscis and wings do not undergo morphological changes.

Keywords: Aedes aegypti, Aedes albopictus, Insect Growth Regulator (IGR), Larvae, Pyriproxyfen.

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#### Introduction

Dengue fever is a world health problem. It is endemic in more than 100 countries and threatens about 2.5 billion or 40% of people living in urban, suburban, and rural tropical and subtropical climates. As of May 2007, dengue fever cases in Indonesia reached 66,365 people with 738 deaths (Kusriastuti, 2007). Various mitigation measures have been undertaken, including vector control utilizing mosquito nest eradication, selective abatization with larvicides, mosquito fogging. In addition to vector control, other efforts were made such as the detection and early treatment of dengue cases (Dinkes Kota Tangerang, 2015).

The primary vector of dengue fever is *Aedes aegypti*, while its secondary vector is *Aedes albopictus* (Djakaria, 1998). According to Sutomo (2003), the vector of dengue fever always lives near residential areas where living conditions and sanitation are inadequate. In Thailand, according to Ponlawat et al. (2005), *Aedes albopictus* is more challenging to control than *Aedes aegypti* because of its more comprehensive range of habitats, found mostly in suburban and rural areas, and in open areas overgrown with plants. *Aedes albopictus* in Thailand has been poorly resistant to temefos, malathion, and permethrin.

Eradication of mosquitoes by using chemicals causes problems such as increased resistance, environmental pollution, poisoning, and nontarget animal deaths. Given the constraints in the use of insecticides, another option to control *Aedes sp.* larvae is needed. The method of insect growth regulator (IGR) is suggested to overcome this problem because it does not affect water cleanliness, does not increase resistance, and it is safe (Munif, 1997). Insecticides used to control the *Aedes sp.* species in water reservoirs should have low toxicity to mammals (WHO, 1995).

Pyriproxyfen is used to control mosquito populations in residential areas, as vector control programs, and as well as to monitor mosquito in husbandry areas. Pyriproxyfen is effective at low doses, low toxicity to mammals, and relatively safe to the environment (Wirawan, 2006). The results of the study of Ali et al. (1995) obtained the results that pyriproxyfen ( $LC_{90} = 0.000376$ ppm) is more toxic 2.23 and 21.5 times compared to diflubenzuron and methoprene. Waris research (2005) about the influence of pyriproxyfen on malaria vector *Anopheles subpictus grassi* showed that there was an abnormality causing mortality in the adult mosquito (larvae and pupae).

Pyriproxyfen has never been applied to dengue fever vectors in Tangerang City. If pyriproxyfen will be used as a means of controlling the dengue fever vector, it is necessary to know the diagnostic concentration for monitoring the mosquito susceptibility in the field. For that purpose, laboratory scale research of the effectiveness of the usage of insect growth regulator (IGR) pyriproxyfen on the growth and the development of *Aedes aegypti* and *Aedes albopictus* larvae in Tangerang City, Indonesia was done with the aim to know the inhibitory power (IE<sub>50</sub> and IE<sub>90</sub>) and LC<sub>50</sub> concentration of pyriproxyfen to *Aedes aegypti* and *Aedes albopictus* larvae.

#### Materials and Methods

The type of research used was laboratory experimental with a post-test design with only one group control design. The subjects were divided into two groups: treatment group and control group. The subjects used were third instar larvae from three sub-district in Tangerang City by installing more than 100 house traps in 100 houses, both inside and outside the home. Determination of the location was based on the endemicity of the region and data on the incidence of dengue cases in recent months.

The treatment was initiated by conducting the preliminary test to determine the variation of  $LC_{50}$  concentration. The pyriproxyfen solution for the preliminary test was prepared as follows: 10g Sumilarv 0.5% (w/w) with active ingredient pyriproxyfen was dissolved in acetone up to 50 mL volume (1000ppm active ingredient). The concentration variations were made by introducing a 0.1ml; 0.2ml; 0.3ml; 0.4ml; 0,5ml; 0.6ml; and 0.7ml pyriproxyfen solution into a test

container containing tap water and 25 instars larvae to obtain the final volume of 200 mL, thus the concentration variations of 0.5ppm; 1.0ppm; 1.5ppm; 2.0ppm; 2.5ppm; 3.0ppm and 3.5ppm were obtained. Observations were made 48 hours after the treatment, and the percentage of larval mortality between 10% to 95% was used as a reference to determine the concentration variation in the actual test by using increment factor. The concentration for larvae mortality test was calculated by increment factor with 6 variations of concentration, i.e. 0.70ppm; 0.98ppm; 1.37ppm; 1.92ppm; 2.69ppm; and 3.76ppm.

The actual test was performed by varying the concentration of 1000 ppm of pyriproxyfen by adding as much as 0.14ml; 0.196ml; 0.274ml; 0.384ml; 0.538ml; and 0.752ml into each test container containing tap water and 25 larvae and fixed to 200 mL volume. The first controls contained 25 larvae in the mixture of 200 mL water and 0.75 2mL acetone and the second one included 25 larvae in 200 mL water. Each treatment was made four replications. Observation of larvae mortality was performed 48 hours after the exposure. The results of inspection of larvae mortality were calculated by probit analysis to obtain the LC<sub>50</sub> value of pyriproxyfen against *Aedes aegypti* and *Aedes albopictus*.

The LC<sub>50</sub> value obtained from probit calculation was replicated four times and used to find the value of LT<sub>50</sub>. The preparation of the stock pyriproxyfen solution was the same as the preliminary test, while the volume addition depends on the LC<sub>50</sub> value. Observations of larvae and pupae deaths were performed every 24 hours. The probit analysis was used to obtain the LT<sub>50</sub> on *Aedes aegypti* and *Aedes albopictus*.

#### Results

Preliminary test results of pyriproxyfen inhibition to the growth and development of third instar larvae at each concentration showed that the pyriproxyfen concentration of 0.05ppb-0.4ppb caused 8%-84% inhibition on the growth of third instar larvae of *Aedes aegypti* and 16%-92% inhibition on the growth of third instar larvae of *Aedes albopictus*. The higher concentration showed the stronger inhibitory effect.

Preliminary test results at the recommended dose of 10ppb; 30ppb; and 50ppb showed that no larvae were successful in maturity, so they were not used in the actual test. The variations of concentration were calculated by increment factor and yielded 0.03ppb; 0.05ppb; 0.08ppb; 0.14ppb; 0.39ppb; and 0.67ppb 0.23ppb; as the concentration used in the actual test. The actual test used third instar larvae with 25 subjects in each treatment and replicated four times so that the total test larvae were 100 subjects in each treatment. The time required for the actual test was nine days at a temperature between 20°C-28°C and humidity between 51% -60%.

On the first day of observation, *Aedes aegypti* larvae had entered the third and fourth instar stage. On day 2, it had started to occur 20%-29% reduction in the number of larvae after partially become pupa. On day 3 of observation, there was some decrease in the number of larvae in each treatment and control, which were varied between 48%-64%. On the 9th day, there was a decrease up to 100% because the larvae had become pupae and adult mosquito. Percentage of the number of pupae per day of observation can be seen in Figure 1 and percentage of the number of adult mosquito can be seen in Figure 2.

#### 60 Percentage of Number of Aedes aegypti 50 Control 40 0,03 Pupae -0,05 30 -0,08 -0,14 20 -0,23 10 0,39 0,67 0 0 1 2 3 4 5 6 7 8 9 Days

**Figure 1** Percentage of number of *Aedes aegypti* pupae on the pyriproxyfen exposure of various concentrations and controls in units of time (days)



**Figure 2** Percentage of number of Adult Mosquito of *Aedes aegypti* on the pyriproxyfen exposure of various concentrations and controls in units of time (days)

On the first day of observation, *Aedes albopictus* larvae have entered the third and fourth in star stage. On day 2, it had started to occur 68%-90% reduction in the number of larvae after partially become pupa. On day 3 of observation, there was some decrease in the number of larvae in each treatment and control, which were varied between

44%-68%. On the 9th day, there was a decrease up to 100% because the larvae had become pupae and adult mosquito. Percentage of the number of pupae per day of observation can be seen in Figure 3 and percentage of the number of adult mosquito can be seen in Figure 4.

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**Figure 3.** Percentage of number of *Aedes albopictus* pupae on the pyriproxyfen exposure of various concentrations and controls in units of time (days)



**Figure 4.** Percentage of number of Adult Mosquito of *Aedes albopictus* on the pyriproxyfen exposure of various concentrations and controls in units of time (days)

The observation of the development of *Aedes aegypti* and *Aedes albopictus* after pyriproxyfen treatment and probit calculation resulted that  $IE_{50}$  value (95% Fiducial limits (FL)) for *Aedes aegypti* was 0.17ppb (0.13-0.21) and  $IE_{90}$  (95% FL) was

0.68ppb (0.43-1.08). IE<sub>50</sub> (95% FL) for *Aedes* albopictus was 0.11ppb (0.09-0.13) and IE<sub>90</sub> (95% FL) was 0.39ppb (0.26-0.57). Inhibitory data on various concentration variations and the result of probit analysis are summarized in Table 1.

**Table 1**  $IE_{50}$  and  $IE_{90}$  value of pyriproxyfen to third instar larvae of *Aedes aegypti* and *Aedes albopictus* after nine days of observation

No	Concentrat ion (ppb)	Number of	Aedes aegypti (n=800)				Aedes albopictus(n=800)	
			%IE	IE <sub>50</sub> (95%FL)	IE <sub>90</sub> (95%FL)	%IE	IE <sub>50</sub> (95%FL)	IE <sub>90</sub> (95%FL)
1	Control	25	0,00			0,00		
2	0,03	25	6,19			8,33		
3	0,05	25	14,43			25,00		
4	0,08	25	23,71	0,17ppb	0,68ppb	39,58	0,11ppb	0,39ppb
5	0,14	25	44,33	(0,13-0,21)	(0,43-1,08)	55,21	(0,09-0,13)	(0,26-0,57)
6	0,23	25	54,64			76,04		
7	0,39	25	82,47			91,67		
8	0,67	25	90,72			97,92		

Preliminary test with pyriproxyfen treatment for the exposure of 48 hours showed the percentage of the death of *Aedes aegypti* at the lowest concentration (0,5ppm) was equal to 8% and at the highest level (3,5ppm) was equal to 84%. The preliminary test of *Aedes albopictus* showed the percentage of death over 48 hours exposure was similar to of 12% at the lowest concentration (0.5ppm) and 88% at the highest level (3.5ppm) as shown in Table 2.

**Table 2.** Mortality percentage of third instar larvae of *Aedes aegypti* and *Aedes albopictus* after 48 hours of exposure to pyriproxyfen in various concentration for the preliminary test

Concentration	Number of	Aedes aegypti		Aedes albopictus	
(nnm)	Number of	No. of Death	Percentage	No. of Death	Percentage
(ppin)	Samples	Larvae	(%)	Larvae	(%)
Control	25	0	0	0	0
0,5	25	2	8	3	12
1,0	25	4	16	5	20
1,5	25	10	40	8	32
2,0	25	14	56	15	60
2,5	25	17	68	18	72
3,0	25	20	80	21	84
3,5	25	21	84	22	88

Effect of concentration variations on *Aedes aegypti* and *Aedes albopictus* larvae resulted in the larvae mortality value that was not much different so that the concentration variations for two species were similar. Variations of concentration to be used to determine  $LC_{50}$  value were based on the results of the calculation of 6 variations, which were: 0.70ppm; 0.98ppm; 1.37ppm; 1.92ppm;

1.37ppm; and 3.76ppm. *Aedes aegypti* mortality data was indicated by  $LC_{50}$  (95% FL) of 1.63ppm (1.38-1.92), and *Aedes albopictus* was indicated by  $LC_{50}$  (95% FL) of 1.56ppm (1.34-1.81). The data of larval mortality on each concentration and the  $LC_{50}$  value of the probit analysis results are summarized in Table 3.

**Table 3**  $LC_{50}$  value of pyriproxyfen to third instar larvae of *Aedes aegypti* and *Aedes albopictus* after 48 hours of exposure

No			Aedes aegypti (n=700)		Aedes albopictus (n=700)	
	Concentration	Number of	Death	LC <sub>50</sub>	Death	$LC_{50}$
	(ppm)	Sample	Percentage	(95%FL)	Percentage	(95%FL)
			(%)		(%)	
1	Control	25	0		1	
2	0,70	25	12		14	
3	0,98	25	25	1 (2	25	1,56ppm (1,34-1,81)
4	1,37	25	40	1,63ppm	37	
5	1,92	25	54	(1,58-1,92)	56	
6	2,69	25	76		84	
7	3,76	25	92		95	

Morphological changes after the treatment of pyriproxyfen were documented. There were noticeable differences between *Aedes aegypti* and *Aedes albopictus* larvae which were dead without



a. Aedes aegypti larvae



c. Aedes aegypti larvae



e. Aedes aegypti larvae

exposure to pyriproxyfen (controls) and those which were exposed to pyriproxyfen. Figure 5 showed the morphological changes of larvae.



b. Aedes albopictus larvae



d. Aedes albopictus larvae



f. Aedes albopictus larvae

Figure 5. a-b. Dead controls larvae

- c-f. Dead larvae after 48 hours exposure to pyriproxyfen
- i. cuticle detached from the thorax and fluid come out from the head
- ii. cuticle disconnected from the thorax and the head swell
- iii. thorax and abdomen damage on segment 1-4

iv. thorax damage and abdomen turned into white-colored

Aedes aegypti and Aedes albopictus pupae exposed to pyriproxyfen gave varying results. In the inhibition test, some of the dead pupae suffered damage to the first and second segment abdomen as shown in Figure 6.c and its entrails came out (Figure 6.d). As a comparison, dead pupae which were not exposed to pyriproxyfen (controls) is shown in Fig. 6.a-b.

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a. Aedes aegypti Pupae





b. Aedes albopictus Pupae



c. Aedes aegypti Pupae
d. Aedes albopictus Pupae
Figure 6. a-b. Dead controls pupae
c-d. Dead pupae treated with pyriproxyfen
i. The body part was damaged in the first and second segment abdomen

ii. Some of the entrails came out from the second abdomen

The results of inhibition tests showed that some mosquitoes could successfully become adults. Observations were made to its legs, probes, and wings. Some of *Aedes aegypti* and *Aedes albopictus* pupae did not succeed to become perfect adult mosquito because the adult mosquitoes could not get out of the pupal case as shown in Figure 7.a-f. Most occurred in the legs that could not be separated from the pupal case, but there was also a wing section that could not get out of the pupal case.



c. Aedes aegypti Mosquito

d. Aedes albopictus Mosquito

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e. *Aedes aegypti* Mosquito f. *Aedes albopictus* Mosquito **Figure 7** a-f. Adult mosquito with some parts of the body that could not get out of its pupal case

- i. Wings and legs could not get out of pupal case
- ii. One of the legs could not get out of pupal case
- iii. Some legs could not get out of pupal case

Some *Aedes aegypti* and *Aedes albopictus* died after reaching adult stage because it could not fly so that its body was always in the water. Some could not operate because the wings could not expand either one or both of



a. Aedes aegypti Mosquito



c. Aedes aegypti Mosquito



e. Aedes aegypti Mosquito





b. Aedes albopictus Mosquito



d. Aedes albopictus Mosquito



f. Aedes albopictus Mosquito

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g. *Aedes aegypti* Mosquito Figure 8. Adult mosquitoes could not fl

h. Aedes albopictus Mosquito

- **Figure 8.** Adult mosquitoes could not fly due to morphological changes in the legs and wings i-ii. Wings could not expand
  - iii-iv. The wings were imperfect, and the straight legs made it was difficult to land on the water surface
  - v. A pair of middle legs curved on each segment
  - vi. The legs were folded because the tip of the tarsus was attached to the abdomen
  - vii. Feet were folded, and wings could not expand
  - viii. Head to abdomen curved

#### Discussion

Exposure to pyriproxyfen slowed down the development of larvae to become pupae and adults. This can be seen from the observation day 9, when it was still found *Aedes aegypti* pupae which lived at concentration 0,05ppb; 0.08ppb; and 0.14ppb respectively 1%; 2% and 1% and *Aedes albopictus* pupa lived at concentrations of 0.03ppb; 0.05ppb; and 0.23ppb respectively by 3%; 1% and 2%.

Inhibition of emergence (IE)<sub>50</sub> (95% FL) of pyriproxyfen against Aedes aegypti and Aedes albopictus were 0.17ppb (0.13-0.21) and 0.11ppb (0.09-0.13) respectively, while IE<sub>90</sub> was 0.68ppb (0.43-1.08) and 0.39ppb (0.26-0.57) respectively. The concentration of pyriproxyfen to inhibit the growth of Aedes aegypti larvae was bigger than Aedes albopictus which showed that Aedes albopictus was more susceptible than Aedes aegypti. Allegedly, this was because of the habit Aedes albopictus that usually live outside the home, so the frequency of exposure to insecticides was less. The difference between the two species was not significant because there was overlap between each confidence limit (FL95%). The response of Aedes aegypti and Aedes albopictus to pyriproxyfen is shown by the regression line with

the equation y = 2,132x + 2,404 and y = 2,445x + 2,480 which can be used to predict the concentration required to obtain the desired percentage of inhibition.

According to Sullivan (2000), pyriproxyfen given at the mosquito larvae stage will function as a juvenile hormone which then binds to the juvenile hormone receptor and will make the larvae cannot develop into an adult. The inhibition test caused many larvae to fail to become live pupae. This was because juvenile hormone disrupted the transformation of late instar larvae into pupae and subsequently matures (WHO, 2005). The failure of pupae to metamorphose into an adult mosquito will cause death in pupa stage (Gaubard, 2007).

The larvae mortality test for determination of  $LC_{50}$  was performed on six series of pyriproxyfen concentration on third instar larvae with the results of  $LC_{50}$  (95% FL) pyriproxyfen against *Aedes aegypti*, and *Aedes albopictus* were 1.63ppm (1.38-1,92) and 1,56ppm (1.34-1.81) respectively. The mosquito response to pyriproxyfen causing death is indicated by the regression line equation y = 3.302x-0.871 for *Aedes aegypti* and y = 3,825x-1,626 for *Aedes albopictus* which can be used to predict the

concentration required to obtain the desired percentage of death.

The larvae mortality test was continued by determining LT<sub>50</sub> at its LC<sub>50</sub> concentration against two species of *Aedes sp.* mosquitoes. The  $LC_{50}$ value of pyriproxyfen against Aedes aegypti was found to be different with Aedes albopictus. The test to search for LT<sub>50</sub> was done at the different concentrations of pyriproxyfen, and the results were also different. The LT<sub>50</sub> values (95% FL) of pyriproxyfen against Aedes aegypti and Aedes albopictus were 57,94 hours (52,25-64,26) and 52,75h (48,43-57,45) respectively. The mosquito response to pyriproxyfen causing death is indicated by the regression line equation y =3.415x + 0.881 for Aedes aegypti and y = 3,734x+ 0,591 for Aedes albopictus which can be used to predict the time required to obtain the desired percentage of death.

Dead larvae indicated the swelling of the thorax. The pupae showed some damages on the cuticle of the first and second abdomen so that there were some organs expelled from the body. Mature mosquitoes that showed three differences. First, perfect adult mosquitoes that can fly and no external morphological change in the legs, wings, and proboscis. Secondly, pupae that become adult mosquitoes but could not get out of the pupal case with some of its organs were still attached to the case. These mosquitoes could survive for several days, and some died. Third, an adult mosquito that could come out of the pupal case but could not fly because there were legs that could not support the body, so it fell in the water. Some were alive when they were removed from the water, but a day later died. It was suspected because of the weak physical, and it could not suck the juice of food in the form of sugar water so that there was a decrease in endurance. Further, it was also found an adult mosquito that could not flap its wings because it was sticky on the thorax or abdomen, so it could not fly. Percentage of those that cannot fly was more significant than the ones that could not get out of the case. According to Tunaz and Uygun (2004), the use of IGR leads to various abnormalities in insects thus decreasing its ability to survive. Siddal (1976) also says a similar thing that IGR caused a variety of defects that disrupt the life of insects although it is not very toxic.

#### Conclusion

Based on the results of the research, it can be concluded as follows:

- 1. *Pyriproxyfen* could inhibit the growth of *Aedes aegypti* larvae with  $IE_{50}$  of 0.17ppb and  $IE_{90}$  of 0.68ppb and inhibit the growth of *Aedes albopictus* larvae with  $IE_{50}$  of 0.11ppb and  $IE_{90}$  of 0.39ppb.
- 2. *Pyriproxyfen* caused death after 48 hours of exposure to *Aedes aegypti* larvae at  $LC_{50}$  of 1.63 ppm and  $LT_{50}$  of 57.94 hours and on *Aedes albopictus* larvae at  $LC_{50}$  of 1.56ppm and  $LT_{50}$  of 52.75 hours.
- 3. External morphological changes occurred in larvae, pupae, and adult mosquitoes, especially on the feet. The wings did not experience morphological changes except the adhesions happened in the abdomen. Proboscis did not change on both *Aedes aegypti* and *Aedes albopictus* larvae.

#### References

- 1. Ali, A., Nayar, J.K., De Xue, R., 1995, Comparative Toxicity of Selected Larvicides and Insect Growth Regulators to A Florida Laboratory Population of Aedes albopictus, Journal of the American Mosquito Control Association, 11(1),72-76.
- Anonim, 2007, Pyriproxyfen(200), http://www.fao org/ag/AGP/AGPP/pesticid /JMPR / /99 eva/25 pyriproxyfen.pdf download tanggal 8 Juli 2007.
- Belding, D.L, 1952, *Textbook of Clinical Parasitology*, Appleton Century Crofts, Inc, New York, 745.
- 4. Chism, B.D, Apperson, C.S., Horizontal transfer of the Insect Growth Regulator pyriproxyfen to larval microcosms by

Gravid Aedes albopictus and Ochloretatus triseriatus Mosquitoes in the Laboratory, Medical and Veterinary Entomology(2003)17:211-220.

- Cutwa M., O'Meara, G. F., 2006, Photographic Guide to Common Mosquitoes of Florida, Florida Medical Entomology Laboratory, University of Florida, Florida.
- Depkes, 2007, Modul Pelatihan Bagi Pengelola Program Pengendalian Penyakit Demam Berdarah Dengue di Indonesia, Ditjen PP PL Depkes, Jakarta.
- Djakaria, S., 1998, Pendahuluan Entomologi dan Vektor Penyakit Virus, Riketsia, Spiroketa, dan Bakteri dalam Parasitologi Kedokteran editor Gandahusada S., Ilahude H. D., Pribadi W., Edisi ketiga, Fakultas Kedokteran Universitas Indonesia, Jakarta, 213.
- Dinkes Kota Tangerang, 2015, Arsip Data DBD di Kota Tangerang.
- 9. Finney, D.J., 1971, *Probit Analysis, Third edition*, At the University Press, Cambridge.
- 10. Gaubard, J., 2007, Juvenile Hormone Binding Proteins Importance in the JH action, Introductory paper no 168, Lund University http://www.kem.ekol. lu.se/introyohann.
- Hoedojo, 1998, Morfologi, Daur Hidup dan perilaku nyamuk dalam Parasitologi Kedokteran editor Gandahusada S., Ilahude H. D., Pribadi W., Edisi ketiga, Fakultas Kedokteran Universitas Indonesia, Jakarta, 221-224.
- Hoedojo, Zulhasriel, 1998, Pengendalian Vektor dalam Parasitologi Kedokteran editor Gandahusada S., Ilahude H. D., Pribadi W., Edisi ketiga, Fakultas Kedokteran Universitas Indonesia, Jakarta, 244-246.
- 13. Kesumawati Hadi,U., Koesharto, 2006,Nyamuk dalam Hama Permukiman Indonesia: Pengenalan, Biologi dan

Pengendalian editor Sigit, S.H, Kesumawati Hadi, U., Institut Pertanian Bogor, Bogor, 23-48

- 14. Kusriastuti, R., 2007, Kebijaksanaan Penanggulangan Demam Berdarah Dengue dalam Simposium Nasional Demam Berdarah Dengue, aspek Biologi Molekuler, Patogenesis, Manajemen dan Pencegahan Kejadian Luar Biasa, Pusat Studi Bioteknologi, Universitas Gadjah Mada, Yogyakarta, 16 Mei 2007.
- 15. Mardihusodo, S.J., 2004, Cara-cara Inovatif Pengamatan dan Pengendalian Nyamuk Vektor, DBD, Pusat Kedokteran Tropis, UGM, Yogyakarta.
- 16. Munif, Amrul, 1997, Pengaruh Residu Pyriproxyfen 0,5% terhadap Larva Aedes aegypti Pada Berbagai Simulasi Wadah Air, Cermin Dunia Kedokteran Nomor 119, Jakarta.
- Ponlawat, A., Scott, J.G., Harrington, L.G., 2005, *Insecticide Susceptibility of Aedes aegypti and Aedes albopictus across Thailand*, J. Medical Entomology, 42(%),821-825.
- Rozendaal, 1997, Vector Control; Methods for Use by Individuals and Communities, WHO, Geneva, 38-39,123.
- 19. Service, M. W., 1996, Medical Entomology for Students, First edition, Chapman&Hall, London.
- 20. Siddal, J.B., 1976, Insect Growth Regulators and Insect Control: A. Critical Appraisal, Environmental Health Persp., Vol 14, 119-126.
- Sihuincha, M., Perea, E.Z., Rioz, W.O., Stancil, J.D., 2005, Potential Use of Pyriproxyfen for Control of Aedes aegypti (Diptera: Culicidae) in Iquitos, Peru, J. Med.Entomol, 42(4), 620-630.
- 22. Sullivan, J., 2000, Environmental Fate of Pyriproxyfen, Environmental Monitoring and Pest Management Branch, Sacramento.

- 23. Sutaryo, 2004, Dengue, Medika Fakultas Kedokteran UGM, Yogyakarta.
- 24. Sutomo, S., 2003, Rencana Strategi Pencegahan &Pemberantasan Demam Dengue/Demam Berdarah Dengue 2004-2006, Ditjen P2M-PL Depkes R.I., Jakarta
- Tunaz, H., Uygun, N., 2004, Insect Growth Regulators for Insect Pest Control, Turk J. Agric For 28(2004), 377-387.
- Waris, L., 2005, Pengaruh pyriproxyfen terhadap vektor malaria Anopheles subpictus grassi asal Pulau Kalimantan dalam Seminar Regional Sulawesi, Palu, 29 September 2005.
- 27. WHO, 1995, Report of the Informal Consultation on Aircraft Disinsection, Geneva.
- 28. WHO, 2001, Report of The Fourth Whopes, Working Group Meeting, Review of IR3535; KBR3023;(RS)-Methoprene 20%EC, Pyriproxyfen 0,5% GR, and Lambda-Cyhalothrin 2,5%CS, WHO/CDC/WHOPES, Geneva, 50.
- 29. WHO, 2005, Guidelines For Laboratory and Field Testing of Mosquito Larvicides, WHO Communicable Disease Control, Prevention, and Eradication, WHOPES, Geneva
- 30. WHO, 2006, *Pyriproxyfen, WHO* specifications and Evaluations for Public Health Pesticides, Geneva.
- 31. Widyana, 1997, Pengaruh Penerapan Metode Pengendalian Vektor Terhadap Populasi Pradewasa Aedes aegypti di Kecamatan Kasihan Bantul, Tesis, UGM, Yogyakarta.